

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES**  
**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

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| <b>Laboratory contact details</b>                                      | Netherlands Institute for Vectors, Invasive plants and Plant health<br>P.O. Box 9102, 6700 HC Wageningen, Netherlands  |
| <b>Short description of the test</b>                                   | This validation data is for generic detection and identification of phytoplasmas. Phytoplasmas can be detected using conventional nested PCR. The conventional (nested) PCR product is purified and finally sequenced using HTS. For identification see validation sheet 571.  |
| <b>Date, reference of the validation report</b>                        | 2022-07-21 - 2020.molbio.012   |
| <b>Link to other validation data</b>                                   | - 2020.molbio.012 This validation data is for generic detection and identification of phytoplasmas. Phytoplasmas can be detected using real time PCR or conventional nested PCR. The conventional (nested) PCR product is purified and finally sequenced using HTS.<br>- 2020.molbio.012 This validation data is for generic detection of phytoplasmas. Phytoplasmas can be detected using real time PCR. For identification see validation sheet 555.<br>- 2020.molbio.004 v1, 2021.molbio.009 v3 This test can be used for the untargeted detection and identification of molecularly characterized ssRNA(+), ssRNA(-), dsRNA, cssRNA, dsDNA(-RT), ssDNA viruses and viroids in symptomatic plant samples. |
| <b>Validation process according to EPPO Standard PM7/98?</b>           | yes  |
| <b>Is the lab accredited for this test?</b>                            | yes  |
| <b>Was the validated data generated in the framework of a project?</b> | no   |
| <b>Description of the test</b>   |  |
| <b>Organism(s)</b>   | 'Candidatus Phytoplasma fragariae' (PHYPPFG)<br>'Candidatus Phytoplasma aurantifolia' (PHYPPAF)<br>'Candidatus Phytoplasma brasiliense' (PHYPPBR)<br>'Candidatus Phytoplasma fraxini' (PHYPPFR)<br>'Candidatus Phytoplasma oryzae' (PHYPPOR)<br>'Candidatus Phytoplasma phoenicium' (PHYPPH)<br>'Candidatus Phytoplasma pruni' (PHYPPN)<br>'Candidatus Phytoplasma pyri' (PHYPPY)<br>'Candidatus Phytoplasma solani' (PHYPPSO)<br>'Candidatus Phytoplasma trifolii' (PHYPPTR)<br>'Candidatus Phytoplasma ulmi' (PHYPPUL)   |

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|   | 'Candidatus Phytoplasma ziziphi' (PHYPZI)<br>Grapevine flavescence dorée phytoplasma (PHYP64)<br>'Candidatus Phytoplasma americanum' (PHYPAE)<br>'Candidatus Phytoplasma palmicola' (PHYPPPL)<br>'Candidatus Phytoplasma palmae' (PHYPPA)<br>Phytoplasma (1PHYPG) |
| <b>Detection / identification</b>   | detection and identification  |
| <b>Method(s)</b>  | Molecular Extraction DNA RNA<br>Molecular Conventional PCR<br>Molecular HTS<br>Molecular other  |
| <b>Method: Molecular Extraction DNA RNA</b>   |   |
| <b>Reference of the test description</b>  |   |
| <b>As or adapted from an EPPO diagnostic protocol</b>   | yes   |
| <b>New test being considered for inclusion in the next version of the EPPO diagnostic protocol?</b> | yes   |
| <b>EPPO Diagnostic Protocol name</b>  | PM 7/133 Generic detection of phytoplasmas (version 1)  |
| <b>As or adapted from an IPPC diagnostic protocol</b>   | no  |
| <b>Is the test modified compared to the reference test</b>  | no  |
| <b>Kit</b>  |   |
| <b>Is a kit used</b>  | yes   |
| <b>Manufacturer name</b>  | QIAGEN  |
| <b>Specify the kit used</b>   | DNeasy Plant Mini Kit   |
| Kit used following the manufacturer's instructions?   | yes   |
| <b>Other information</b>  |   |
| <b>Method: Molecular Conventional PCR</b>   |   |
| <b>Reference of the test description</b>  |   |
| <b>As or adapted from an EPPO diagnostic protocol</b>   | yes   |
| <b>New test being considered for inclusion in the next version of the EPPO diagnostic protocol?</b> | no  |
| <b>EPPO Diagnostic Protocol name</b>  | PM 7/133 Generic detection of phytoplasmas (version 1)  |
| <b>Name of the test</b>   | Conventional nested PCR using the primers P1/P7 and R16F2n/R16R2  |
| <b>As or adapted from an IPPC diagnostic protocol</b>   | yes   |
| <b>IPPC diagnostic Protocol name</b>  | ISPM 27 Annex 12 DP 12: Phytoplasmas (version   |

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|   | 2016)  |
| <b>Name of the test</b>   | Conventional nested PCR  |
| <b>Is the test modified compared to the reference test</b>  | no   |
| <b>Kit</b>  |  |
| <b>Is a kit used</b>  | no   |
| <b>Other information</b>  |  |
| <b>Reaction type</b>  | Nested   |
| <b>Method: Molecular HTS</b>  |  |
| <b>Reference of the test description</b>  |  |
| <b>As or adapted from an EPPO diagnostic protocol</b>   | no   |
| <b>New test being considered for inclusion in the next version of the EPPO diagnostic protocol?</b> | no   |
| <b>As or adapted from an IPPC diagnostic protocol</b>   | no   |
| <b>Reference of the test</b>  | Addendum - New Supporting information for PM 7/151 Considerations for the use of high throughput sequencing in plant health diagnostics, appendix 1.   |
| <b>Is the test modified compared to the reference test</b>  | no   |
| <b>Other information</b>  |  |
| <b>Other details on the test</b>  | The same pipeline for viruses and viroids is used to detect phytoplasmas. See data validation 512 and Addendum - New Supporting information for PM 7/151 Considerations for the use of high throughput sequencing in plant health diagnostics, appendix 1. |
| <b>Method: Molecular other</b>  |  |
| <b>Reference of the test description</b>  |  |
| <b>As or adapted from an EPPO diagnostic protocol</b>   | no   |
| <b>New test being considered for inclusion in the next version of the EPPO diagnostic protocol?</b> | no   |
| <b>As or adapted from an IPPC diagnostic protocol</b>   | no   |
| <b>Reference of the test</b>  | QIAquick PCR Purification Kit (Qiagen)   |
| <b>Is the test modified compared to the reference test</b>  | no   |
| <b>Kit</b>  |  |
| <b>Is a kit used</b>  | yes  |
| <b>Manufacturer name</b>  | QIAGEN   |
| <b>Specify the kit used</b>   | QIAquick PCR Purification Kit (Qiagen)   |

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| Kit used following the manufacturer's instructions?                     | yes  |
| <b>Other information</b>  |  |
| <b>Other details on the test</b>  | Purification of PCR-product before sequencing (HTS)  |
| <b>Performance Criteria :</b>   |  |
| <b>Organism 1.:</b>   | 'Candidatus Phytoplasma fragariae'(PHYPPG)   |
| <b>Organism 2.:</b>   | 'Candidatus Phytoplasma aurantifolia'(PHYPAF)  |
| <b>Organism 3.:</b>   | 'Candidatus Phytoplasma brasiliense'(PHYPPB)   |
| <b>Organism 4.:</b>   | 'Candidatus Phytoplasma fraxini'(PHYPPF)   |
| <b>Organism 5.:</b>   | 'Candidatus Phytoplasma oryzae'(PHYPOR)  |
| <b>Organism 6.:</b>   | 'Candidatus Phytoplasma phoenicium'(PHYPPH)  |
| <b>Organism 7.:</b>   | 'Candidatus Phytoplasma pruni'(PHYPPN)   |
| <b>Organism 8.:</b>   | 'Candidatus Phytoplasma pyri'(PHYPPY)  |
| <b>Organism 9.:</b>   | 'Candidatus Phytoplasma solani'(PHYPSO)  |
| <b>Organism 10.:</b>  | 'Candidatus Phytoplasma trifolii'(PHYPTR)  |
| <b>Organism 11.:</b>  | 'Candidatus Phytoplasma ulmi'(PHYFUL)  |
| <b>Organism 12.:</b>  | 'Candidatus Phytoplasma ziziphi'(PHYZZI)   |
| <b>Organism 13.:</b>  | Grapevine flavescence dorée phytoplasma(PHY64)   |
| <b>Organism 14.:</b>  | 'Candidatus Phytoplasma americanum'(PHYPAE)  |
| <b>Organism 15.:</b>  | 'Candidatus Phytoplasma palmicola'(PHYPPAL)  |
| <b>Organism 16.:</b>  | 'Candidatus Phytoplasma palmae'(PHYPPA)  |
| <b>Organism 17.:</b>  | Phytoplasma(1PHYPP)  |
| <b>Analytical sensitivity</b>   |  |
| <b>What is smallest amount of target that can be detected reliably?</b> | Grapevine flavescence doree phytoplasma is detected at 10 <sup>3</sup> dilution in the real time PCR (Hodgetts et al. 2009). GFDP is detected at 10 <sup>2</sup> dilution in the conventional nested PCR (Lee et al., 1993 and Gundersen & Lee, 1996).   |
| <b>Analytical specificity - inclusivity</b>                             |  |
| <b>Number of strains/populations of target organisms tested</b>         | see annex 4 (validation report) and table 1 in (addendum). Ca. Phytoplasma aurantifolia Ca. Phytoplasma aurantifolia-related strain SPLL Ca. Phytoplasma brasiliense Ca. Phytoplasma fraxini-reference strain Ca. Phytoplasma oryzae Ca. Phytoplasma phoenicium Ca. Phytoplasma pruni Ca. Phytoplasma pyri Ca. Phytoplasma solani Ca. Phytoplasma trifolii Ca. Phytoplasma ulmi Ca. Phytoplasma ulmi Ca. Phytoplasma ziziphi Grapevine flavescence doree phytoplasma |

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|  | Grapevine flavescence doree phytoplasma GFDP<br>Map-FD1 Grapevine flavescence doree<br>phytoplasma GFDP Map-FD2 Grapevine flavescence<br>doree phytoplasma GFDP Map-FD2 PEY05<br>Grapevine flavescence doree phytoplasma GFDP<br>Map-FD3 Ca. Phytoplasma americanum Ca.<br>Phytoplasma palmae Ca. Phytoplasma palmicola |
| <b>Specificity value</b>   | 100   |
| <b>Reproducibility</b>   |   |
| <b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b> | 100   |
| <b>Repeatability</b>   |   |
| <b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b> | 100   |
| <b>Test performance study</b>  |   |
| <b>Test performance study?</b>   | no  |
| <b>Other information</b>   |   |
| <b>Any other information considered useful</b>   | see PM 7/079 (2) for grapevine flavescence doree<br>phytoplasma.  |
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| The following complementary files are available<br>online:                               | <ul style="list-style-type: none"> <li>• <a href="#">Validation report</a></li> <li>• <a href="#">Validation report annex</a></li> <li>• <a href="#">Validation report addendum</a></li> <li>• <a href="#">2020.molbio.012 Detection and identification of Candidatus Phytoplasma fragariae V1.0</a></li> </ul>         |

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