

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES**  
**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Laboratory contact details</b>	Anses Plant Health Laboratory - Bacteriology, Virology and GMO Unit 7 rue Jean Dixméras, 49044 Angers, France
<b>Short description of the test</b>	detection of Xylella fastidiosa Xylella fastidiosa by Molecular real time PCR in Specimen
<b>Date, reference of the validation report</b>	2020-03-04 - MA065 version 01 report version 1
<b>Validation process according to EPPO Standard PM7/98?</b>	yes
<b>Is the lab accredited for this test?</b>	yes
<b>Was the validated data generated in the framework of a project?</b>	Euphresco
<b>If yes, please specify</b>	Euphresco project 09/2016-08/2018 "Harmonized protocol for monitoring and detection of Xylella fastidiosa in its host plants and its vectors" AND H2020 Ponte project 2016-2020
<b>Description of the test</b>	
<b>Organism(s)</b>	Xylella fastidiosa (XYLEFA)
<b>Detection / identification</b>	detection
<b>Method(s)</b>	Molecular Extraction DNA RNA Molecular real time PCR
<b>Method: Molecular Extraction DNA RNA</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an EPPO diagnostic protocol</b>	yes
<b>New test being considered for inclusion in the next version of the EPPO diagnostic protocol?</b>	yes
<b>EPPO Diagnostic Protocol name</b>	PM 7/024 Xylella fastidiosa (version 4)
<b>As or adapted from an IPPC diagnostic protocol</b>	no
<b>Is the test modified compared to the reference test</b>	yes Analyse on single specimen and on composite sample pooling till 10 specimens
<b>Kit</b>	
<b>Is a kit used</b>	yes

<b>Manufacturer name</b>	BIONOBILE
<b>Specify the kit used</b>	QuickPick™ SML Plant DNA
Kit used following the manufacturer's instructions?	yes
<b>Other information</b>	
<b>Method: Molecular real time PCR</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an EPPO diagnostic protocol</b>	yes
<b>New test being considered for inclusion in the next version of the EPPO diagnostic protocol?</b>	no
<b>EPPO Diagnostic Protocol name</b>	PM 7/024 Xylella fastidiosa (version 4)
<b>Name of the test</b>	Real-time PCR - duplex (Harper et al., 2010; erratum 2013)
<b>As or adapted from an IPPC diagnostic protocol</b>	no
<b>Is the test modified compared to the reference test</b>	no
<b>Kit</b>	
<b>Is a kit used</b>	no
<b>Other information</b>	
<b>Reaction type</b>	Duplex - Probe
<b>Other details on the test</b>	A cut-off value of 38 is applied
<b>Performance Criteria :</b>	
<b>Organism 1.:</b>	<b>Xylella fastidiosa(XYLEFA)</b>
<b><u>Analytical sensitivity</u></b>	
<b>What is smallest amount of target that can be detected reliably?</b>	1.10 <sup>3</sup> cells in a single insect and in composite sample grouping till 10 insects with a detection rate of 100% and with a cut-off value of 38 Ct
<b><u>Diagnostic sensitivity</u></b>	
<b>Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98</b>	Not evaluated
<b><u>Analytical specificity - inclusivity</u></b>	
<b>Number of strains/populations of target organisms tested</b>	14 target strains Cf. attached file "Rapport de validation MA065 v1"
<b>Specificity value</b>	100%
<b><u>Analytical specificity - exclusivity</u></b>	
<b>Number of non-target organisms tested</b>	40 non target organisms Cf. attached file "Rapport de validation MA065 v1"
<b>Specificity value</b>	100%
<b><u>Diagnostic Specificity</u></b>	

<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>	Not evaluated
<b><u>Reproducibility</u></b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% at $10^3$ cells/head Single sample (one head) : Evaluated with 3 DNA extract replicates per concentration and 3 amplification replicates par DNA extracts by 1 operators on 3 different days. 3 bacterial concentration tested : $10^2$ cells/mL; $10^3$ cells/mL, $10^4$ cells/mL and healthy samples. Composite sample (10 heads) : Evaluated with 3 DNA extract replicates per concentration and 3 amplification replicates par DNA extracts by 1 operators on 2 different days. 3 bacterial concentration tested : $10^3$ cells/mL; $10^4$ cells/mL, $10^5$ cells/mL and healthy samples. Bacterial concentrations evaluated by ddPCR.
<b><u>Repeatability</u></b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% at $10^3$ cells/head Single sample (one head) : Evaluated with 3 DNA extract replicates per concentration and 3 amplification replicates par DNA extracts by 1 operators on 3 different days. 3 bacterial concentration tested : $10^2$ cells/mL; $10^3$ cells/mL, $10^4$ cells/mL and healthy samples. Composite sample (10 heads) : Evaluated with 3 DNA extract replicates per concentration and 3 amplification replicates par DNA extracts by 1 operators on 2 different days. 3 bacterial concentration tested : $10^3$ cells/mL; $10^4$ cells/mL, $10^5$ cells/mL and healthy samples. Bacterial concentrations evaluated by ddPCR.
<b>Test performance study</b>	
<b>Test performance study?</b>	yes
<b>Brief details of the test performance study and its output. It available, link to published article/report</b>	Reference: 17-XFAST-EU (Euphresco / H2020 Ponte project) TPS organized in 2017 by Anses. For the present method, 5 laboratories participated. Samples were constituted by Phileanus spumarius head (one head : sample) spiked with a Xf bacterial suspension in order to get final concentrations of $5.10^3$ to $5.10^5$ cells/head. Performance criteria Results Sensitivity 100% Specificity 100% Accuracy 100% Repeatability 99% Reproducibility 100% Limit of detection (with detection rate of 100%) $5.10^3$ cells/head of insect
<b>Other information</b>	
<b>Any other information considered useful</b>	Bibliography : Cuntz C., Legendre B., De Jerphanion P., Juteau V., Forveille A., Germain J-F., Ramel J-M., Reynaud P., Olivier V. et Poliakoff F., (2020), Xylella fastidiosa subspecies and sequence types detected in Philaenus spumarius and in infected plants in France share the same locations, Plant Pathology, 00 : 1-14.

The following complementary files are available online:

- [Rapport de validation MA065 v1](#)

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